



Galdieria-Medium

Original recipe as in:

Gross & Schnarrenberger (1995), modified according to Allen's *Cyanidium* Medium, modified (Allen 1959, Watanabe *et al.* 2000).

further references:

Allen, M.B. (1959): Studies with *Cyanidium caldarium*, an anomalously pigmented chlorophyte. - *Arch. Mikrobiol.* **32**(3): 270-277.

Gross, W. & Schnarrenberger, C. (1995): Heterotrophic growth of two strains of the acido-thermophilic red alga *Galdieria sulphuraria*. - *Plant Cell Physiol.* **36**(4): 633-638.

Watanabe et al. 2000: > see present online-catalogue of NIES collection

For 1000 mL final culture medium add the following quantities (Volume/Mass) of stock solutions (SL) prepared at the given concentrations to 850 mL dd-H₂O. Add <u>one component after the other until</u> <u>each one has completely mixed</u> and finally fill up to 1000 mL.

All stock solutions can be stored unsterilised at 4 °C. Store sterile-filtered vitamin mix (SL 12) at -20 °C.

Stock Solution (SL)	Volume or Mass	Component	Concentration in SL	Concentration in final Medium
- - - -	1.50 g 0.30 g 0.30 g 1 mL	(NH₄)2SO₄ MgSO₄ • 7H₂O KH₂PO₄ CaCl₂• 2H₂O	- - 2 g • 100 mL ⁻¹	1.1 • 10 ⁻² M 1.2 • 10 ⁻³ M 2.2 • 10 ⁻³ M 1.3 • 10 ⁻⁴ M
Fe-EDTA	2.07 mL	EDTA (not as Na-salt) FeSO4 ∙ 7H₂O KOH 1n	0.52 g • 100 mL ⁻¹ 0.5 g • 100 mL ⁻¹ 5.4 ml • 100 mL ⁻¹	3.66 • 10 ⁻⁵ M 3.70 • 10 ⁻⁵ M 1.11 • 10 ⁻⁶ M
Trace elements solution (Allen metals)	1 mL	$\begin{array}{l} ZnSO_{4} \cdot 7H_{2}O \\ H_{3}BO_{3} \\ CoCL_{2} \cdot 6H_{2}O \\ CuSO_{4} \cdot 5H_{2}O \\ (NH_{4})_{6}Mo_{7}O_{27} \cdot 6H_{2}O \\ NaVO_{3} \\ MnCl_{2} \cdot 4H_{2}O \end{array}$	220 mg • 1 L ⁻¹ 2860 mg • 1 L ⁻¹ 40 mg • 1 L ⁻¹ 79 mg • 1 L ⁻¹ 130 mg • 1 L ⁻¹ 40 mg • 1 L ⁻¹ 1790 mg • 1 L ⁻¹	7.65 • 10 ⁻⁷ M 4.63 • 10 ⁻⁵ M 1.68 • 10 ⁻⁷ M 6.40 • 10 ⁻⁶ M 2.10 • 10 ⁻⁷ M 3.28 • 10 ⁻⁷ M 9.04 • 10 ⁻⁶ M

Combine all trace elements in one SL. Dissolve each component completely one after the other. It may need autoclaving to dissolve. Trace elements solution should **not** be stored in glass containers, but instead in teflon or polycarbonate containers to prevent adsorption of metals to container surface.

Adjust medium to final pH of 1.8 or as desired with 0.5 M H_2SO_4 and autoclave at 121 $^\circ C$ for 20 min.

For stock cultures on agar slants add 1.0-1.3 % Agar (e.g. purified high strength, 1000 g \cdot cm⁻²) to prepared medium before autoclaving. The pH must be lowered to 4-4.5, otherwise the agar will not solidify.

1 % glucose may be added for heterotrophic cultivation in the dark.